Review Article

6

Resveratrol as a potential protective compound against metabolic inflammation

Hossein Ghahremani¹, Arash Bahramzadeh², Kosar Bolandnazar³, Solaleh Emamgholipor², Hossein Hosseini⁴, Reza Meshkani^{2*}

¹ Department of Clinical Biochemistry, Faculty of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran

² Department of Clinical Biochemistry, Faculty of Medicine, Tehran University of Medical Sciences, Tehran, Iran

³ Department of Biological Sciences and Technology, Islamic Azad University of Mashhad, Mashhad, Iran

⁴ Department of Clinical Biochemistry, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

Article info:

Received: 12 June 2023 Revised: 18 July 2023 Accepted: 27 July 2023

<u>ABSTRACT</u>

Although the exact mechanism linking obesity to type 2 diabetes (T2D) remains unknown, accumulating evidence suggests that low-grade chronic metabolic inflammation or 'metainflammation' plays a pivotal role. Adipose tissue is the primary site of meta-inflammation, and overproduction of pro-inflammatory cytokines in this tissue affects other organs such as the liver, skeletal muscle, pancreas, and brain. This leads to the development of insulin resistance and metabolic irregularities in these tissues. Therefore, strategies targeting metainflammation could be effective in treating T2D and related metabolic traits. Resveratrol, a polyphenol, is suggested to possess anti-inflammatory and immunomodulatory activities. The anti-inflammatory effect of resveratrol is mediated through several mechanisms including the suppression of nuclear factor kB (NF-kB), down-regulation of extracellular signal-regulated kinase (ERK)/p38 mitogen-activated protein kinase (MAPK) signaling, suppression of toll-like receptor (TLR)-mediated pathway; inhibition of NLR family pyrin domain containing 3 (NLRP3) inflammasome activation, reduction of reactive oxygen species (ROS) generation, suppression of immune cell infiltration into tissues; and inhibition of pro-inflammatory cytokines production. This review will examine the evidence on the role of resveratrol in modulating inflammation in various organs affected by obesity such as liver, skeletal muscle, kidney, heart and brain

Keywords: Resveratrol, metabolic inflammation, type 2 diabetes, macrophage, Adipose tissue

Abbreviations: AP1: activator protein 1; ATMs: adipose tissue macrophages; AMPK: AMPactivated protein kinase; CVD: cardiovascular disease; CNS: central nervous system; CCR: chemokine (C-C motif) receptor ; CKD: chronic kidney disease; COX: cyclooxygenase ; ERK: extracellular signal-regulated kinase; FFA: free fatty acid; HFD: high fat diet; hs-CRP: high sensitive C reactive protein; JNK: Jun N-terminal kinase; ICAM-1: intercellular adhesion molecule-1; IFN- γ : interferon γ ; IL: interleukin; IKK β : inhibitor of κ B kinase; I/R: ischemic/ reperfusion; LPS: lipopolysacharide; MIP-1 α : macrophage inflammatory protein 1 α ; MCP-1: macrophage chemoattractant protein-1; MAPK: mitogen-activated protein kinase; NKT: natural killer T cells; NO: nitric oxide; iNOS: nitric oxide synthase; NAFLD: non-alcoholic fatty liver disease; NASH: non-alcoholic steatohepatitis; NF-kB: nuclear factor kB; Nrf-2: nuclear factor erythroid 2-related factor 2; NLRP3: NLR family pyrin domain containing 3; PARP: poly (ADP-ribose) polymerase; ROS: reactive oxygen species; SIRT1: sirtuin 1; SVF: stromavascular fraction; SOCS3: suppressor of cytokine signaling-3; TLR: toll-like receptor; T2D: type 2 diabetes; TNF- α : tumor necrosis factor α ; TGF β : transforming growth factor β ; VCAM-1: vascular cell adhesion molecule-1; WAT: white adipose tissue

Use your device to scan and read the article online

* Corresponding Author:

Department of Clinical Biochemistry, Faculty of Medicine, Tehran University

of Medical Sciences, Tehran, Iran

Email: rmeshkani@tums.ac.ir

Reza Meshkani,

Citation: Ghahremani H, Bahramzadeh A, Bolandnazar K, Emamgholipor S, Hosseini H, Meshkani R. Resveratrol as a potential protective compound against metabolic inflammation. Acta Biochimica Iranica. 2023;1(2):50-64.

doi https://doi.org/10.18502/abi.v1i2.14101



Copyright © 2023 Tehran University of Medical Sciences. Published by Tehran University of Medical Sciences

This work is licensed under a Creative Commons Attribution-NonCommercial 4.0 International license(https://creativecommons.org/licenses/by-nc/4.0/) Noncommercial uses of the work are permitted, provided the original work is properly cited.

Introduction



ccording to a report by the World Health Organization (WHO), overweight and obesity are major risk factors for several chronic diseases, including type 2 diabetes (T2D), cardiovascular

diseases (CVD), and cancer (1). A systematic analysis for the Global Burden of Disease Study revealed that the prevalence of overweight and obesity in adolescents reached 36.9% in men and 38% in women in 2013 (2).

Obesity is known to be associated with low-grade chronic inflammation (3). This inflammation induced by obesity is closely linked to insulin resistance, T2D, atherosclerosis, and non-alcoholic fatty liver disease (NAFLD) (4). This low-grade inflammation, known as meta-inflammation, is characterized by increased levels of circulating pro-inflammatory cytokines and acute phase proteins, as well as enhanced recruitment of immune cells, especially macrophages, to adipose tissue. The cytokines and chemokines produced by macrophages cause local and systemic inflammation, which in turn leads to pancreatic β -cell dysfunction and peripheral insulin resistance (5).

In recent years, the use of natural plant products for the prevention/treatment of various chronic inflammatory diseases has attracted increased attention (6-9). Among many bioactive molecules derived from plants, polyphenols are of particular interest due to their potential anti-inflammatory effects (10). Over the last few decades, increasing evidence has focused on the field of metabolic inflammation to illustrate the beneficial effect of anti-inflammatory agents in the treatment of T2D and its complications. Resveratrol, due to its protective roles in inflammatory responses, is of particular interest for ameliorating meta-inflammation associated diseases. This review will summarize the effects of resveratrol on ameliorating meta-inflammation in different metabolic tissues.

Metabolic inflammation

By definition, metabolic inflammation or metainflammation is considered a chronic low-grade inflammation, triggered by a set of metabolic and inflammatory cells in response to an overload of nutrients or other intrinsic cues (11, 12). This definition differs from classical inflammation, which is the acute reaction of the immune system to cope with injuries and infection and is characterized by redness, swelling, fever, pain, heat, and edema (11, 13).

Adipose tissue is recognized as the central tissue in meta-inflammation, and overproduction of cytokines in this tissue might affect several other organs such as the liver, skeletal muscle, pancreas, and brain (11, 14). Apart from the secretion of inflammatory cytokines, adipocytes in obese subjects contribute to activating inflammatory processes through recruiting inflammatory

cells such as macrophages and lymphocytes. Metainflammation can be activated by multiple factors such as inflammatory cytokines, excess free fatty acids (FFAs), reactive oxygen species (ROS), hypoxia and endoplasmic reticulum (ER) stress, inflammasome activation and alteration in gut microbiota (11). Notably, meta-inflammation is considered the central hallmark of all metabolic consequences of obesity such as insulin resistance, T2D and atherosclerosis (5).

Adipose tissue is considered the main source of inflammatory cytokines in T2D. Structurally, adipose tissue is composed of white adipose tissue (WAT), and brown adipose tissue. WAT is suggested to be the main central part of adipose tissue involved in metainflammation. WAT is further divided into subcutaneous and visceral fat tissues, with distinct physiology and function in pathologic processes. Abdominal WAT seems to play a major role in meta-inflammation (15). The mature adipose cells and stroma-vascular fraction (SVF) constitute two central parts of WAT (16). SVF itself is composed of extracellular matrix with dispersed fibroblasts, preadipocytes, endothelial, and immune cells such as macrophages, CD3⁺ T cells (CD4⁺ or CD8⁺), mast cells, eosinophils and B cells. Adipocytes are the major cell population in the adipose tissue; however, the SVF is functionally important for this tissue (17). It is generally accepted that excessive fat accumulation can alter both the number and function of immune cells. Specifically, fat accumulation increases the number and activity of macrophages, mast cells, neutrophils, T- and B lymphocytes but reduces eosinophils and several subclasses of T lymphocytes including Treg, T helper 2 (Th2) and natural killer T (NKT) cells. This imbalance plays a significant role in the development of obesityrelated local and systemic inflammation.

The increased infiltration of immune cells is considered the main characteristic of adipose tissue inflammation (18, 19). Evidence from rodent and human studies reveals that macrophages constitute more than 40% of the total adipose tissue content of obese subjects, compared to almost 10% in lean counterparts (20). Based on cytokine profile and surface markers, adipose tissue macrophages (ATMs) are categorized into two main subsets: M1 and M2. In lean conditions, adipocytes produce several cytokines, such as interleukin (IL)-13, that induce activation of M2 macrophages. Alternatively activated macrophages (M2) secrete anti-inflammatory factors, such as IL-10 and transforming growth factor β (TGF- β) (18, 20). Classically activated macrophages (M1) secrete pro-inflammatory cytokines such as tumor necrosis factor α (TNF- α) and IL-6. Obesity can cause a switch from the M2 to M1 phenotype, characterized by a decrease in the production of anti-inflammatory cytokines and an increase in the production of proinflammatory cytokines. Adipocyte hypertrophy induced by over-nutrition promotes hypoxia-mediated death of adipocytes. Crown-like structures are formed as a result

of macrophage accumulation in hypoxic regions around the dead adipocytes (18). These events coincide with the release of several pro-inflammatory cytokines and chemokines, resulting in a vicious cycle amplifying ATM content and propagating the chronic inflammatory state. Indeed, pro-inflammatory cytokines and macrophagesecreted factors themselves exert paracrine effects to activate inflammatory pathways within insulin target cells. This leads to activation of Jun N-terminal kinase (JNK), inhibitor of κB kinase β (IKK β), and other serine kinases that subsequently activate transcription factor targets, including activator protein 1 (AP1) and nuclear factor-kB (NF-kB). Serine kinases also phosphorylate insulin receptor substrate proteins, insulin receptors, and possibly other insulin signaling molecules, thereby interrupting the normal function of insulin and leading to insulin resistance (20).

In addition to macrophages, other leukocyte populations can infiltrate adipose tissue. Obesity is characterized by a remarkable elevation of CD8⁺ T cells and a decrease in both CD4⁺ helper and regulatory T cells (Treg) in visceral adipose tissue. CD8+ T cells are involved in the differentiation, recruitment, and activation of macrophages during obesity. However, CD4⁺ T helper and Treg cells play a fundamental role in determining the outcome of inflammatory responses. It is well known that TH1 cells are pro-inflammatory cells and enhance macrophage secretion of inflammatory cytokines, while TH2 and Treg cells induce an anti-inflammatory M2 macrophage phenotype. NKT cells have been observed in obese adipose tissue and recognize lipid antigens. Depletion of NKT cells decreases insulin resistance and adipose tissue inflammation following high-fat diet (HFD) feeding, while activation of these cells exacerbates metabolic effects of diet-induced obesity (21).

Mast cells accumulate in obese adipose tissue earlier than macrophages and produce and secrete various mediators, such as histamine, serotonin, heparin, serine protease, eicosanoids, and cytokines. Among these mediators, IL-6 and interferon γ (IFN- γ) have a notable impact on inflammation associated with obesity (18, 21). These cytokines increase protease expression in adipose tissue, which in turn leads to angiogenesis by degrading anti-angiogenic molecules and further infiltration of leukocytes into the adipose tissue (21).

Dendritic cells are specialized antigen-presenting leukocytes involved in macrophage activation and accumulation (M1) at the site of inflammation (18). Dendritic cells express a number of surface markers such as CD11c, which are similar to the markers used to characterize the M1 subpopulation of adipose tissue macrophages in mice (18). CD11c is a member of the β 2-integrins and has been used as an activation marker for monocytes/macrophages (22-23).

Neutrophils are the first immune cells recruited to inflamed tissues and can enhance the consequent infiltration of inflammatory monocytes by secreting monocyte chemoattractant protein-1 (MCP-1) and other chemokines. The IL-4 and IL-13 induce the M2polarisation of macrophages in WAT (24). The levels of eosinophils, one of the main sources of IL-4 and IL-13 in adipose tissue, are reduced during obesity. Eosinophils improve glucose homeostasis by sustaining the levels of anti-inflammatory M2 macrophages in the WAT, possibly through an IL-4 and IL-13 signaling-dependent manner (18).

As stated earlier, meta-inflammation has been suggested as the underlying mediator of obesity-induced T2D. Therefore, it is reasonable to conclude that strategies targeting meta-inflammation could be interesting options for treatment and prevention of metabolic disorders. In vitro and in vivo evidence has suggested that resveratrol could be a promising therapeutic agent against metabolic inflammation. In the following sections, this review will discuss in more detail the effects of resveratrol on metabolic inflammation in different tissues.

Resveratrol

In recent years, understanding the "French Paradox" has drawn increasing attention to investigate whether polyphenolic antioxidants such as resveratrol may be useful beyond the cardiovascular system. The French paradox refers to the paradoxical association of a diet high in saturated fat and cholesterol with low CVD mortality, which was initially attributed to consumption of red wine and its constituent resveratrol (25).

Resveratrol, chemically known as 3,5,4'-trihydroxystilbene, is a naturally occurring nonflavonoid polyphenolic antioxidant compound produced in a large variety of plant species such as mulberries, peanuts, and grapes (26-28). Resveratrol was first isolated from the roots of white hellebore (Veratrum grandiflorum O. Loes) in 1940, and later, in 1963, from the roots of Polygonum Cuspidatum (29). Its name has been derived from its source; a resorcinol derivative coming from a Veratrum species (30). Phenylalanine is a precursor of resveratrol biosynthesis that, following several reactions, is converted to 4-coumaroyl-CoA. Finally, the enzyme stilbene synthase converts one molecule of p-coumaroyl-CoA and three molecules of malonyl-CoA into resveratrol. There is evidence that biosynthesis of resveratrol is induced in plants in response to physical and chemical stresses (31). Resveratrol is a photosensitive molecule susceptible to isomerization via UV radiation. This polyphenol exists naturally as both cis- and trans-isomers; however, the trans-isomer is believed to be the most abundant and biologically active form (32).

There is increasing evidence that resveratrol prevents or attenuates progression of a variety of disorders, as well as improves stress resistance and extends the lifespans of various organisms from yeast to vertebrates (29). Since the discovery of caloric restriction-like effects of resveratrol, it was shown that resveratrol might provide beneficial effects against CVD, metabolic diseases, and cancer. Based on in vitro and in vivo studies, the beneficial effect of resveratrol is mediated through its anti-aging, anti-carcinogenic, anti-inflammatory, and anti-oxidant activities (33).

Resveratrol and diabetes

There is increasing evidence showing the beneficial effects of resveratrol on alleviating insulin resistance and T2D (32). Clinical trials in T2D patients point to the antidiabetic effects of resveratrol (26, 34). More recently, a meta-analysis of 11 randomized controlled trials demonstrated that resveratrol consumption significantly improves insulin sensitivity and glycemic control in patients with diabetes (35). Moreover, data from rodent studies have indicated the effects of resveratrol on improving insulin sensitivity, decreasing hyperglycemia, enhancing insulin secretion, and enhancing antioxidant capacity in pancreatic β -cells (36). In addition, a decrease in HbA1c concentrations in an animal model of diabetes (streptozotocin-nicotinamide-induced diabetic rats) after resveratrol administration reveals the anti-diabetic potential of this natural polyphenol (37).

Several mechanisms, including a decrease in oxidative stress, reduction of inflammation, down-regulation of protein-tyrosine phosphatase 1B, prevention of pancreatic β cell loss, and decrease in lipid accumulation in muscle and liver tissues, have been suggested to explain the anti-diabetic effects of resveratrol (33, 37). Moreover, resveratrol enhances the lipolytic response to epinephrine and reduces the insulin ability to counteract lipolysis in adipose tissue. Resveratrol also reduces hyperglycemia in diabetic rats (32). The underlying mechanisms for resveratrol benefits in glucose homeostasis are induction of sirtuin 1 (SIRT1) activity, stimulation of glucose uptake by increased expression and membrane trafficking of GLUT4, activation of glucose uptake in the absence of insulin, elevation of glycogen synthase activity and decrease of glycogen phosphorylase activity in the liver (32, 37). However, other mechanisms including the activation of 5'-AMPactivated protein kinase (AMPK) play an important role in induction of fatty acid oxidation (32).

Resveratrol as an anti-inflammatory agent

There is accumulating evidence that resveratrol, as an anti-inflammatory agent, inhibits the expression and secretion of pro-inflammatory mediators such as TNF- α , IL-6, IL-1, IL-12, and IFN- γ . Recently, a metaanalysis of 17 randomized controlled trials revealed that resveratrol consumption significantly reduces serum and plasma levels of TNF- α and high sensitive C reactive protein (hs-CRP) (38). Additionally, the inhibitory role of resveratrol in eicosanoid synthesis and activation of several enzymes such as inducible nitric oxide synthase (iNOS), cyclooxygenase-1 (COX-1), or COX-2 has also been reported (39, 40). It has been shown that the anti-inflammatory property of resveratrol is mediated through the down-regulation of transcription factors NF-kB or AP-1 (27).

Resveratrol and inflammation in adipose tissue

As mentioned above, adipose tissue is a central organ in meta-inflammation. In this section, the effects of resveratrol on the inflammatory processes in adipose tissue are described. Several studies have provided data that resveratrol functions as an anti-inflammatory agent in adipose tissue and exerts its pivotal effects on reversing the deleterious actions of obesity on inflammation in T2D. The anti-inflammatory effect of resveratrol on adipose tissue has been studied in both immune cells and adipocytes.

Studies have reported the effect of resveratrol on macrophage infiltration into adipose tissue. It has been suggested that this polyphenol markedly attenuates visceral WAT inflammation and insulin resistance in a mouse model of sleep apnea. In detail, sleep fragmentation augments tissue inflammation and insulin resistance through recruiting macrophages to visceral WAT, reducing M2 polarity cell counts, suppressing the proportion of circulating Treg cells, and increasing the population of M1 polarity in mice. Interestingly, resveratrol treatment reversed all these alterations (41). In another study, Jeon et al. demonstrated that resveratrol reduced macrophage migration to adipose tissue in mice fed a HFD (42). In addition, intermittent hypoxia caused an increase in the total number of macrophages in visceral WAT which consists of an increase in the pro-inflammatory M1 macrophage and reduction in M2 macrophages. These changes were markedly abrogated in resveratrol administration state (43). Nøhr et al. reported that resveratrol supplementation alleviated lipopolysaccharide (LPS)-induced inflammation in mice which was associated with a reduced expression of TNF- α and IL-1 β in epididymal fat. Resveratrol was not able to reduce macrophage infiltration, but it switched their phenotype into a more anti-inflammatory state (M2 macrophage) in the epididymal adipose tissue (44). Moreover, Lv Zm et al. evaluated macrophage infiltration in fat depots by measuring crown-like structure (CLS) density, which consisted of dead adipocytes and infiltrating macrophages. Histological analysis showed that a high-calorie and high-cholesterol diet increased adipocyte size and the density of CLSs in mice and resveratrol supplementation prevented these alterations (45). Resveratrol also reduced proinflammatory cytokines via suppressing macrophage infiltration and inhibiting NF-KB activity in adipose tissue of genetically obese rats (46).

There is accumulating evidence regarding the effects of resveratrol on adipocytes. In human adipose tissue explants, resveratrol reversed IL-1 β -induced inhibition of adiponectin production (47). Moreover,

resveratrol suppressed ROS production and forskolininduced lipolysis in human differentiated adipocytes (48). There is also a report that oral administration of resveratrol ameliorated adipose tissue dysfunction in fructose-fed rats through inhibiting NF-kB p65 phosphorylation, abrogating the expression of the adipocytokines, pro-inflammatory and elevating adiponectin and peroxisome proliferator-activated receptor (PPAR) expressions in perivascular adipose tissue in an AMPK/SIRT1-interdependent manner (49). Chronic administration of resveratrol improved insulin sensitivity and inflammatory responses in visceral WAT of diet-induced obese monkeys and 3T3-L1 adipocytes possibly through upregulating SIRT1 expression, decreasing adipocyte size, and suppressing NF-kB activation. Remarkably, serum from resveratroltreated monkeys suppressed NF-kB activation as well as expression of IL-6 and IL-1 β in 3T3-L1 adipocytes (50). In addition, SIRT1 activation by resveratrol diminished the expression of CD40 in 3T3-L1 adipocytes-treated with TNF- α partially via modulating NF- κ B-dependent pathway (51). Further experiments have also shown that resveratrol suppresses obesity-associated inflammatory through downregulation of plasminogen activator inhibitor 1 (PAI-1) gene expression in vitro model of inflamed adipose tissue possibly through suppressing NF- κ B activity. It was also demonstrated that signaling via phosphatidyl inositol 3 kinase (PI3K), SIRT1, AMPK, ROS, and nuclear factor erythroid 2-related factor 2 (Nrf-2) were not involved in mediating the inducing effects of resveratrol on PAI-1 production (52). In addition, it is evident that hypoxia and fibrosis are early triggers of the adipose dysfunction in obesity. Resveratrol effectively ameliorated fibrosis and inflammation in adipose tissue which was associated with HIF-1a degradation in a SIRT1-dependent manner (53). Importantly, resveratrol suppressed up-regulation of the inflammatory cytokines such as TNF- α , IFN- α and IFN- β , and their upstream signaling molecules, including toll-like receptor 2 (TLR2), TLR4, MyD88, Tirap, TRAF6, IRF5, TRIF, p-IRF3, and NF-kB in epididymal adipose tissues of obese mice (54). Moreover, resveratrol inhibited TNF-a-induced IL-6, PAI-1, MCP-1 (55, 56) and IL-8 levels in adipocytes. This effect was mainly due to the NF-kB inhibitory potential of resveratrol (24, 57, 58). Specifically, this polyphenol can repress NF-kB binding activity directly or indirectly through the activation of SIRT1 (58). Other potential mechanisms for the antiinflammatory activity of resveratrol includes suppression of the activation of extracellular signal-regulated kinase (ERK), c-June N-terminal kinase (JNK), c-Jun, and also activation of PPARy activity (59). Resveratrol also up-regulated the expression of adiponectin, PPAR-α, PPAR-y, SIRT1 and AMPK in epididymal fat depots of HFD-induced obese mice (60). The expression of adipose triglyceride lipase (ATGL) and its downstream PPAR α -mediated lipid signaling pathway was reduced

in both an animal model of aging and aged 3T3-L1 adipocytes, which coincided with upregulation of TNF- α and IL-6 production. Notably, resveratrol attenuated the production of pro-inflammatory cytokines in an ATGL/PPARα-dependent manner (61). Resveratrol also inhibited visceral adipogenesis through suppression of the galanin-mediated signaling molecules such as E2F1, and p-ERK and key adipogenic genes such as fatty acid synthase (FAS) and lipoprotein lipase in HFDinduced obesity. Furthermore, it attenuated cytokine production in adipose tissue by repressing TLR2 and TLR4-mediated inflammation (54). Resveratrol also maintained mitochondrial integrity by inhibiting Drp1 activity and prevented NLR family pyrin domain containing 3 (NLRP3) inflammasome activation by suppressing endoplasmic reticulum (ER) stress. These events were associated with reduced cell apoptosis in the adipose tissue of diabetic mice (62).

Resveratrol and inflammation in liver

Chronic inflammation is the hallmark of insulin resistance in the liver (63, 64). There is compelling evidence to support a central role for systemic and hepatic inflammation in the pathogenesis of non-alcoholic fatty liver disease (NAFLD) and non-alcoholic steatohepatitis Preventing hepatic meta-inflammation (NASH). through diet, exercise, lifestyle modifications, and pharmacological intervention might be helpful in the treatment of NAFLD and insulin resistance (65). In this regard, chronic supplementation of resveratrol in NAFLD patients significantly decreased inflammatory markers (hs-CRP, IL-6, and NF-kB) and cytokeratin-18 (as a biomarker of hepatocellular apoptosis) (66). In another study, resveratrol significantly increased adiponectin levels and reduced the levels of TNF- α , and fibroblast growth factor 21 (FGF-21) in patients with NAFLD (67).

Macrophages are widely distributed in the liver and they are a promising target for resveratrol intervention. Macrophages in the liver are classified into two subsets: (CD68+) Kupffer cells which are engaged in phagocytic activity and (CD11b+) Kupffer cells with cytokine producing capacity (TNF-a, IL-12), that are involved in inflammation and antitumor immunity. Resveratrol was shown to increase the number of phagocytic (CD68+) Kupffer cells and subsequently facilitated uptake of the lipid droplets by these cells. Moreover, resveratrol reduced the capacity of CD11b+ Kupffer cells to produce TNF- α (68). Resveratrol also conferred resistance to hepatocyte steatosis and apoptosis in an animal model of NAFLD which coincided with a significant increase in M2 Kupffer cells polarization (69). Resveratrol administration suppressed Kupffer cells recruitment and down-regulated the expression of pro-inflammatory cytokines such as TNF-α and IL-6 in bile duct ligation and CCL4-induced liver injuries models (70, 71). Furthermore, resveratrol dramatically

inhibited inflammation in a low-dose LPS-induced model of NASH (72). In contrast to above findings, Jeong et al. by measuring macrophage infiltration marker (F4/80), M1 macrophage marker (CD11c) and M2 macrophage marker (CD163), reported that resveratrol treatment could not improve obesity-induced macrophage infiltration and phenotypic switching from M1 to M2 state in the liver of HFD mice (73).

At the molecular level, several targets have been proposed for the anti-inflammatory effects of resveratrol. It was found that resveratrol could relieve HFDinduced inflammation through upregulation of hepatic expression of both SIRT1 and SIRT6. Furthermore, the beneficial effects of resveratrol on glycemic control and NAFLD were accompanied by alterations in NLRP3 inflammasome. Notably, resveratrol administration could reverse increased expression of NLRP3 components including ASC, CASP-1, NALP-1 and NALP-3 in old mice liver (65). Resveratrol prevented HFD-induced hepatic steatosis and ER stress by regulation of the expression of PPAR-δ, ATP citrate lyase, suppressor of cytokine signaling-3 (SOCS3), TNF- α , and IL-1 β (74). Furthermore, administration of resveratrol remarkably inhibited inflammatory responses by suppressing NF-kB p65 expression and reducing the IL-1ß cytokine level in the hepatic tissues of streptozotocin-induced type 1 diabetic rats (75). It has also been suggested that the inhibitory effect of resveratrol on activity of the NF-kB may be mediated by the restoration of its inhibitor; Ik-Ba (76). COX2, a marker of pro-inflammatory innate immune activity, was also upregulated in aged liver and reversed by resveratrol in old mice liver (77). There is ample evidence about the close link between oxidative stress and inflammation in the pathogenesis of liver injury. In this regard, resveratrol significantly reduced the hepatic expression of 4-hydroxynonenal; a marker of oxidative stress in mice with cholesteric liver injury (70).

Resveratrol and inflammation in pancreas

Impaired insulin secretion due to either β -cell dysfunction and/or β -cell loss is now recognized in the pathogenesis and progression of T2D (78). T2D is associated with increased islet-associated immune cells. Indeed, an increased number of islet-associated macrophages was observed early in HFD mice, the *db/db* mouse, and in type 2 diabetic patients during disease progression. Both the exposure of cultured islets to a type 2 diabetic milieu and islet isolation from HFD fed mice were associated with increased levels of islet-derived inflammatory factors including IL-6, IL-8, granulocyte colony-stimulating factor, and macrophage inflammatory protein 1 α (MIP-1 α) (79).

There is evidence regarding resveratrol effects on inflammatory cytokines and inflammatory signaling pathways in pancreatic tissue. It was reported that resveratrol decreased infiltration of neutrophil cells into the pancreas in severe acute pancreatitis (SAP) (80). Oral administration of dihydro-resveratrol also decreased production of TNF- α and activity of NF-kB in the pancreas of cerulein-treated rats (81). Furthermore, resveratrol administration significantly suppressed chemokine (C-C motif) receptor (CCR) 6 production in immune cells and down-regulated CCR6 expression in Th17 and CD11b+F4/80hi macrophages in a nonobese mouse model of type 1 diabetes. Resveratrol also reduced IFN- γ expression and its suppressive functions in T helper (Th)17 polarization. These effects were associated with inhibition of macrophage migration from peripheral lymphoid organs to the pancreas in the NOD mouse model of type 1 diabetes (82).

Resveratrol and inflammation in skeletal muscle

Skeletal muscle is another major site of insulin resistance in obesity and T2D. Skeletal muscle is the primary site of glucose disposal and accounts for almost 80% of insulin-stimulated glucose uptake in the body and, therefore, muscle insulin resistance has a profound effect on glucose intolerance and hyperglycemia in obesity and T2D (5, 83). Studies have suggested that lipid accumulation inside muscle cells, activation of the inflammatory signals in myocytes following exposure to stimuli such as LPS and FFAs, and recruitment of macrophages are some proposed mechanisms underlying skeletal muscle insulin resistance (5, 13). A high level of inflammatory markers such as TNF-a, IL-6, and IL-18 and increased infiltration of macrophages has been reported from skeletal muscle tissue of obese and insulin-resistant subjects (84).

The beneficial anti-inflammatory effects of resveratrol in skeletal muscle cells have been reported in several studies. In support of this notion, microarray analysis on vastus lateralis muscle biopsies revealed that 30 days of resveratrol supplementation in healthy obese subjects reduced the gene expression of genes involved in inflammation (85). Furthermore, resveratrol treatment decreased induction of iNOS, and nitric oxide (NO) production in skeletal muscle of LPS-challenged mice. In vitro studies revealed that the inhibitory effect of resveratrol on both iNOS protein induction and NO production in cytokine/LPS-treated L6 myocytes is partly mediated through a mechanism involving the activation of AMPK (86). Findings also demonstrated that resveratrol has an anti-inflammatory function in skeletal muscle C2C12 cells. Resveratrol pretreatment robustly attenuated palmitate-induced TNF-α and IL-6 expression in C2C12 cells. Molecular analyses revealed that the beneficial effects of resveratrol were accompanied by inhibiting ROS production and decreasing the activity of the MAPKs and NF-kB signaling pathways (87). In contrast to above reports, other studies have shown no effect of resveratrol on skeletal muscle inflammation. It was suggested that resveratrol did not affect the mRNA and protein levels of TNF- α and iNOS in skeletal muscle



Figure 1. Possible effects of resveratrol on modulating inflammation in various organs affected by obesity. At the molecular level, several mechanisms have been proposed for resveratrol effects in ameliorating meta-inflammation. These mechanisms include suppression of infiltration of immune cells into tissues, inhibition of pro-inflammatory cytokines production, inhibition of NLRP3 inflammasome activation, oxidative stress suppression and SIRT1 activation. The modulation of these processes by resveratrol is associated with alleviating the meta-inflammation-induced tissue damage in liver, pancreas, skeletal muscle, kidney, heart and brain

AMPK: AMP-activated protein kinase; CFB: complement factor B; COX-2: Cyclooxygenase-2; GLUT4: Glucose transporter type 4; HIF-1α: Hypoxia-inducible factor 1-alpha; hs-CRP: A high-sensitivity c--reactive protein; ICAM-1: Intercellular Adhesion Molecule 1; IFNα: Interferon-alpha; IFNβ: Interferon-beta; IL-10: Interleukin 10;IL-1β: Interleukin 1beta;IL-6: Interleukin 6; iNOS: Inducible nitric oxide synthase; MCP-1: Monocyte chemoattractant protein-1; NLRP3: NLR Family Pyrin Domain Containing 3; Nrf-2: Nuclear factor (erythroid-derived 2)-like 2; PAI-1: Plasminogen activator inhibitor-1; PGC-1α: Peroxisome proliferator-activated receptor gamma coactivator 1-alpha; ROS: Reactive oxygen species;

TGF-β1: Transforming growth factor beta 1; TNF-α: Tumor necrosis factor alpha

of aged human subjects. Furthermore, 8 weeks of daily intake of resveratrol in aged human subjects did not affect the abundance of the I κ B- α and I κ B- β or the JNK, p65, p38, and IKK phosphorylation (88). In addition, resveratrol supplementation did not alter the expression of TNF- α , IL-6, and MCP-1 in skeletal muscle of HFDinduced obese mice (89). In another report, resveratrol had no effect on CD14 (a marker of macrophage infiltration), TNF- α , and IL1- β expression in skeletal muscle of LPS-treated mice (44). Altogether, the results from above studies suggest that further studies are still needed to provide greater insight into the effects of resveratrol on the inflammatory responses in skeletal muscle cells.

Resveratrol and inflammation in kidney

It is well-accepted that obesity-induced inflammation plays a significant role in the initiation or development of chronic kidney diseases (CKD) and glomerulopathy (90, 91). Indeed, insulin resistance and inflammation are common features in CKD patients (92). Several in vivo and in vitro studies have indicated that infiltration of inflammatory cells (93-97) and elevation of inflammatory cytokines (98-102), play a significant role in the pathogenesis of renal injuries.

Resveratrol, through different signaling alterations, exhibits renoprotective effects in various animal models. In rat renal mesangial cells, resveratrol prevented high glucose-induced cell proliferation and fibronectin expression through inhibiting both NF-κB/NADPH oxidase pathway and ROS production (103). Palsamy et al. demonstrated the beneficial effect of resveratrol on nephropathy in a model of streptozotocin-nicotinamideinduced diabetic rats. Oral administration of resveratrol to diabetic rats significantly normalized the levels of TNF-α, IL-1β, IL-6, NO and NF-κB p65 subunit in renal tissues (104). In another study, it was revealed that resveratrol treatment inhibited renal lipotoxicity, oxidative stress and inflammation by enhancing the AMPK-SIRT1-PGC1α signaling pathway in db/db mice (105). The administration of resveratrol significantly ameliorated LPS-induced acute kidney injury (AKI) in mice by reducing infiltrating cells and also inflammatory cytokine levels in kidney tissue (45). Interestingly, the suppressive effects of resveratrol on infiltration of macrophages and neutrophils have been reported in animal models of cystic kidneys, ischemic/reperfusion (I/R)-induced renal injury and sepsis-induced renal injury (108-110). Treatment of the AKI model with resveratrol also induced apoptosis in macrophages through downregulation of iNOS, Bcl-2, and Bcl-xL and decreased inflammation by suppressing TLR-4 activation and cytokine release (111). Moreover, resveratrol attenuated polycystic kidney disease (PKD) progression by inhibitory role in production of MCP-1, and TNF- α possibly via a pathway dependent on NF-kB (108).

In vitro and animal studies demonstrate a beneficial anti-inflammatory role for resveratrol in different chronic kidney diseases. There is no direct clinical evidence showing the protective and anti-inflammatory effects of resveratrol in CKD patients, however, a few studies imply its beneficial effects on human subjects. In this regard, dietary supplementation with red grape juice exerted anti-oxidative and anti-inflammatory efficacies in hemodialysis patients (112, 113). However, further investigations are required to reveal the potential beneficial effects of resveratrol in human kidney injuries.

Resveratrol and inflammation in heart

The increased incidence of CVD will lead to an expected worldwide number of CVD-related deaths of more than 23.6 million by 2030 (114, 115). Resveratrol has shown beneficial and protective effects against most degenerative and CVD, including I/R injury, atherosclerosis, hypertension, and heart failure.

There is accumulating evidence to support the central role of inflammation in the process of atherosclerosis. Notably, inflammation is involved in all stages of atherosclerosis including initiation, progression and plaque formation (116). Both in vivo and in vitro antiinflammatory effects of resveratrol in atherosclerosis and the underlying mechanism have been suggested. Consistent with this, resveratrol treatment has been shown to inhibit IL-6 and IL-8 production in human coronary artery smooth muscle cells (117), inhibit IL-6 release by stimulated macrophages (118), and reduce serum levels of IL-1 β , IL-6, and TNF- α in an atherosclerotic rabbit model (119). Resveratrol effects on NF-kB, an important transcription factor regulating various mediators or inflammation including cytokines, adhesion molecules, and growth factors was reported (120). Resveratrol has been shown to reduce the expression of adhesion molecules intercellular adhesion molecule-1 (ICAM-1), and vascular cell adhesion molecule-1 (VCAM-1) via inhibiting the NF-κB pathway (121). Similarly, resveratrol suppressed hyperglycemia-induced ICAM-1 expression in endothelial cells through inhibiting the p38 mitogen-activated protein kinase (MAPK) pathway

(122). In cultured endothelial cells, the beneficial effects of resveratrol on inflammatory angiogenesis have been attributed to inhibition of COX-2 and matrix metalloproteinase-9 (MMP-9) (123). Resveratrol also counteracted oxysterol-induced inflammation and NF-kB activation in human macrophages, suggesting the potential effects of resveratrol in preventing or treating atherosclerosis (124). Furthermore, resveratrol has been shown to reverse the pro-inflammatory phenotype in vascular smooth muscle cell of aged non-human primates, suggesting a vasoprotective effect of resveratrol in animal models of aging (125). Resveratrol effects on inflammation could be mediated by modulating the expression of miRNAs that can be anti-inflammatory (e.g., miR-663) or pro-inflammatory (e.g., miR-155) (126).

Inflammation is also a key player in myocardial ischemia. TNF- α can trigger the inflammatory reaction caused by myocardial I/R. Vascular endothelial cell injury, and inflammatory cells, such as neutrophils activated by cytokines, and adhesion molecules are also involved in the inflammatory responses (127). In a recent study, rats with myocardial I/R injury were administered with resveratrol at the onset of reperfusion. The results indicated the propagation of the cascade of inflammatory responses through activating NLRP3 inflammasome and secreting the inflammatory cytokines IL-1ß and IL-18. The authors showed that resveratrol may exert protective effects against I/R injury by inhibiting the expression and activation of the NLRP3 inflammasome (128). Moreover, resveratrol suppressed the inflammatory reaction in rat heart with myocardial I/R by inhibiting TLR4/NF-KB signaling, activating the Nrf2/ARE pathway, increasing NO production and inhibiting both neutrophil infiltration and TNF α expression (129).

Heart failure is a clinical syndrome that arises from a variety of pathophysiologies including hypertension, myocardial infarction, and congenital cardiomyopathies (130). Data from animal studies have reported that the administration of resveratrol prevents and/or slows the progression of heart failure in animal models of heart failure induced by myocardial infarction, pressure overload, and myocarditis (29). Gupta et al. demonstrated that resveratrol treatment significantly attenuated adverse cardiac remodeling in pressure overload-induced heart failure in C57/BL6 mice. In this study, oxidative stress, left ventricular macrophages and mast cells infiltration was significantly increased after 4 weeks of transverse aortic constriction compared to sham-operated mice. Treatment with resveratrol significantly abolished this inflammatory response when compared to transverse aortic constriction alone. This response has been shown to be mediated, at least in part, by inhibition of oxidative stress and inflammation (131). Resveratrol significantly decreased cellular infiltration, fibrosis, and expression of inflammatory cytokines in the myocardium in myosininduced autoimmune myocarditis of rats (132).

Despite the promise of resveratrol as a potential therapeutic for numerous CVD, the number of clinical studies investigating the cardioprotective effects of resveratrol is still limited. Moreover, there is still a great deal of uncertainty on the beneficial effects of resveratrol on inflammatory markers in patients with CVD. In one study, one-year daily intake of a resveratrol-enriched grape extract modulated the expression of key proinflammatory cytokines and inflammation-related microRNAs in peripheral blood mononuclear cells of T2D and hypertensive patients with coronary artery disease (133). In addition, chronic daily consumption of a resveratrol-rich grape supplement improved the inflammatory status in patients at high CVD risk (134, 135). This effect was mediated by increasing adiponectin (136, 137) and IL-10 levels, and suppressing several well-known inflammatory mediators including TNF-a, IL-6/IL-10 ratio, PAI-1, hs-CRP, soluble ICAM-1 (137), and IL-6 (135). On the other hand, receiving 10 mg resveratrol capsule daily for 3 months did not alter CRP and TNF- α in patients with stable coronary artery disease. However, it improved left ventricle diastolic function and endothelial function and reduced LDLcholesterol level (138). In summary, preclinical studies using resveratrol have demonstrated that this polyphenol can ameliorate inflammatory state in CVD, but there are some controversies about the clinical studies, thus there is a need for large and well-controlled clinical trials in this regard.

Resveratrol and inflammation in central nervous system

Inflammation is a complex responsive cascade in the central nervous system (CNS) by which it defends against potentially injurious stimuli. An increasing body of literature has demonstrated that activated microglia and infiltrating monocytes, along with inflammatory cytokines secreted from these cells, have an important role in the initiation and progression of neuro-inflammation (139, 140). According to several studies, resveratrol has been considered to ameliorate neuro-inflammation in the CNS (141, 142).

Firstly, resveratrol as an inhibitor of neuro-inflammation has been investigated in several animal model studies (143-147). For instance, resveratrol had a decreasing effect on the levels of inflammatory factors in the hippocampus of kindled mice and rats (148, 149). It has been reported that the antidepressant function of resveratrol was partially mediated by decreasing the levels of pro-inflammatory cytokines through the NF- κ B signaling pathway in the brain areas of animal models (150, 151). Secondly, there are also several cell models of neuro-inflammation such as CNS-resident microglia, BV-2 murine microglia, N9 microglia and CNSinfiltrating peripheral macrophages, which have been used in studies investigating the cellular mechanisms underlying the anti-inflammatory effects of resveratrol in the CNS (152-155). Moreover, astrocytes as the most abundant cells in the CNS and producers of proinflammatory cytokines (156, 157) can also be affected by resveratrol (158).

There is also evidence that HFD and obesity can result in high levels of FFAs and cytokines in blood (159-161). Following the arrival of FFAs, cytokines and immune cells in the hypothalamus, neuro-inflammation is triggered which in turn leads to interruption of satiety signals and adverse impacts on cognition (162). Although the exact mechanisms by which obesity induces neuro-inflammation is still unclear, there is evidence for intracellular disturbances and stresses such as ER stress, oxidative stress, and autophagic defects in this regard (163). Moreover, diabetic neuropathy is one of the common complications of diabetes and clinical manifestations such as numbness, burning and tingling sensation and intractable pain are attributed to this disorder. The pathophysiological factors of diabetic neuropathy may include hyperglycemia, oxidative stress, lipid peroxidation, nitrosative stress, poly (ADPribose) polymerase (PARP) over activation, defective neurotrophism and autoimmune-mediated nerve destruction which can trigger neuro-inflammation. Following this condition, permanent nerve damage can happen through decreasing nerve functionality and nerve blood perfusion (164, 165).

It has been described that resveratrol attenuated hypothalamic inflammation through a decrease in mRNA levels of IkB and IKKB and protein level and acetylated NF-kB subunit RelA/p65 in a mouse model of T2D (166). Furthermore, resveratrol reduced TNF- α levels in hippocampus of HFD fed diabetic mice (42). It was also described that the Jak-Stat pathway as an activator of the pro-inflammatory factors including IL-15 and IL-22 showed lower expression following resveratrol supplementation in hippocampus of diabetic mice (167). In addition, DNA fragmentation, oxidative stress and levels of malondialdehyde, peroxynitrite, NF-KB, TNF-α, IL-6 and COX-2 indicated a significant decrease following resveratrol treatment in diabetic rats (168, 169). Another study reported that the neuroprotective effect of resveratrol was enhanced in combination with 4-amino 1,8 naphthalimide (4-ANI) as an PARP inhibitor in experimental diabetic neuropathy (170).

A number of studies have reported the possible role of resveratrol in protecting against brain damage following ischemia, stroke, seizure, and epilepsy (171). Rats treated with resveratrol indicated increased heme oxygenase neuroprotective enzyme and decreased intracellular heme levels as a pro-oxidant following stroke (172). Moreover, resveratrol as a cardio- and neuro-protector ameliorated blood-brain barrier disruption and brain damage following recurrent stroke through activation of SIRT-1 and attenuating the inflammatory and oxidative stress markers in adult rats (173). Additionally, resveratrol decreased the volume of infarction following focal cerebral ischemia in rats (174). As discussed above, based on animal models and cell culture studies, it seems that resveratrol mitigates meta-inflammation pertaining to several CNS conditions. However, these results should be confirmed in human studies.

Conclusion

Existing evidence supports the role of low-grade chronic inflammation or metabolic inflammation as a probable mechanism connecting obesity and T2D. Multiple mechanisms including inflammatory cytokines, macrophage infiltration into tissues, excessive FFAs, production of ROS, hypoxia and ER stress, inflammasome activation and alteration in gut microbiota have been implicated in triggering meta-inflammation. Therefore, it is worth mentioning that strategies targeting meta-inflammation could be promising approaches for the treatment and prevention of obesity-related disorders.

Resveratrol, due to its protective roles in the regulation of inflammatory responses, is of particular interest for ameliorating diseases linked with meta-inflammation. The specific mechanisms linked to resveratrol in the context of meta-inflammation are still uncertain, but several mechanisms have been attributed to the anti-inflammatory activities of this polyphenol. These mechanisms include (1) suppression of NFkB signaling pathway by either inhibiting NF-kB p65 phosphorylation or by repressing NF-kB binding activity or by restoration of its inhibitor; Ik-Ba; (2) downregulation of ERK/p38 MAPK signaling pathway; (3) suppression of TLR-mediated pathways; (4) inhibition of NLRP3 inflammasome activation; (5) inhibition of ROS production; (6) suppression of infiltration of immune cells into tissues; and (7) inhibition of proinflammatory cytokines (e.g., TNF-a, IL-1β, IL-6, and CRP) production. The modulation of these processes by resveratrol is associated with protective effects against meta-inflammation-induced tissue injury in adipose tissue, liver, pancreas, skeletal muscle, kidney, heart and brain.

In the case of human studies, the anti-inflammatory properties of resveratrol have been reported in several pre-clinical studies; however, some studies have reported opposing results. The conflicting data may be related to the poor bioavailability of resveratrol in humans and dissimilarities in study design and use of different doses of resveratrol. Therefore, long-term follow-up studies are needed to generalize the beneficial effects of resveratrol in meta-inflammation-related abnormalities to humans..

Conflict of Interest

The authors declare that they have no conflict of interest.

References

- 1. World Health Organization. Obesity: preventing and managing the global epidemic. Geneva: World Health Organization; 2000.
- Ng M, Fleming T, Robinson M, Thomson B, Graetz N, Margono C, et al. Global, regional, and national prevalence of overweight and obesity in children and adults during 1980-2013: a systematic analysis for the Global Burden of Disease Study 2013. Lancet. 2014;384(9945):766-81.
- Rosa FT, Zulet MÁ, Marchini JS, Martínez JA. Bioactive compounds with effects on inflammation markers in humans. Int J Food Sci Nutr. 2012;63(6):749-65.
- Ferrante A. Obesity-induced inflammation: a metabolic dialogue in the language of inflammation. J Intern Med. 2007;262(4):408-14.
- Meshkani R, Vakili S. Tissue resident macrophages: Key players in the pathogenesis of type 2 diabetes and its complications. Clin Chim Acta. 2016.
- Bisht K, Wagner KH, Bulmer AC. Curcumin, resveratrol and flavonoids as anti-inflammatory, cyto-and DNA-protective dietary compounds. Toxicology. 2010;278(1):88-100.
- Zamani-Garmsiri F, Emamgholipour S, Rahmani Fard S, Ghasempour G, Jahangard Ahvazi R, Meshkani R. Polyphenols: potential anti-inflammatory agents for treatment of metabolic disorders. Phytother Res. 2022;36(1):415-32.
- Tehrani SS, Goodarzi G, Panahi G, Zamani-Garmsiri F, Meshkani R. The combination of metformin with morin alleviates hepatic steatosis via modulating hepatic lipid metabolism, hepatic inflammation, brown adipose tissue thermogenesis, and white adipose tissue browning in highfat diet-fed mice. Life Sci. 2023;323:121706.
- Goodarzi G, Tehrani SS, Panahi G, Bahramzadeh A, Meshkani R. Combination therapy of metformin and p-coumaric acid mitigates metabolic dysfunction associated with obesity and nonalcoholic fatty liver disease in high-fat diet obese C57BL/6 mice. J Nutr Biochem. 2023;118:109369.
- Leiherer A, Mündlein A, Drexel H. Phytochemicals and their impact on adipose tissue inflammation and diabetes. Vascul Pharmacol. 2013;58(1):3-20.
- Dali-Youcef N, Mecili M, Ricci R, Andrès E. Metabolic inflammation: connecting obesity and insulin resistance. Ann Med. 2013;45(3):242-53.
- Lumeng CN, Saltiel AR. Inflammatory links between obesity and metabolic disease. J Clin Invest. 2011;121(6):2111-7.
- Khodabandehloo H, Gorgani-Firuzjaee S, Panahi G, Meshkani R. Molecular and cellular mechanisms linking inflammation to insulin resistance and β-cell dysfunction. Transl Res. 2016;167(1):228-56.
- Gerner RR, Wieser V, Moschen AR, Tilg H. Metabolic inflammation: role of cytokines in the crosstalk between adipose tissue and liver 1. Can J Physiol Pharmacol. 2013;91(11):867-72.
- Lontchi-Yimagou E, Sobngwi E, Matsha TE, Kengne AP. Diabetes mellitus and inflammation. Curr Diab Rep. 2013;13(3):435-44.
- Liu L, Mei M, Yang S, Li Q. Roles of chronic low-grade inflammation in the development of ectopic fat deposition. Mediators Inflamm. 2014;2014.
- 17. Mraz M, Haluzik M. The role of adipose tissue immune

cells in obesity and low-grade inflammation. J Endocrinol. 2014;222(3):R113-R127.

- Pereira SS, Alvarez-Leite JI. Low-Grade Inflammation, Obesity, and Diabetes. Curr Obes Rep. 2014;3(4):422-31.
- Jahangard R, Tehrani SS, Emamgholipour S, Vatannejad A, Meshkani R. Autophagy protects peripheral blood mononuclear cells from high glucose-induced inflammation and apoptosis. Acta Biochim Iranica. 2023;1(1):40-9.
- Olefsky JM, Glass CK. Macrophages, inflammation, and insulin resistance. Annu Rev Physiol. 2010;72:219-46.
- Anderson EK, Gutierrez DA, Hasty AH. Adipose tissue recruitment of leukocytes. Curr Opin Lipidol. 2010;21(3):172.
- Chen Y, Tian J, Tian X, Tang X, Rui K, Tong J, et al. Adipose tissue dendritic cells enhances inflammation by prompting the generation of Th17 cells. PLoS One. 2014;9(3):e92450.
- Taghizadeh N, M S, Saeedi V, Haghighi L, Nourbakhsh M, Nourbakhsh M, et al. Association between Steroid Hormones and Insulin Resistance in Patients with Polycystic Ovary Syndrome. Acta Biochim Iranica. 2023;1:1.
- Chmelar J, Chung KJ, Chavakis T. The role of innate immune cells in obese adipose tissue inflammation and development of insulin resistance. Thromb Haemost. 2013;109(3):399-406.
- 25. Artaud-Wild SM, Connor S, Sexton G, Connor W. Differences in coronary mortality can be explained by differences in cholesterol and saturated fat intakes in 40 countries but not in France and Finland. A paradox. Circulation. 1993;88(6):2771-9.
- de Ligt M, Timmers S, Schrauwen P. Resveratrol and obesity: can resveratrol relieve metabolic disturbances? Biochim Biophys Acta Mol Basis Dis. 2015;1852(6):1137-44.
- Kulkarni SS, Cantó C. The molecular targets of resveratrol. Biochim Biophys Acta Mol Basis Dis. 2015;1852(6):1114-23.
- Gorgani-Firuzjaee Sattar, Meshkani Reza. Resveratrol reduces high glucose-induced de-novo lipogenesis through mTOR mediated induction of autophagy in HepG2 cells. Acta Biochim Iranica. 2023;1(1):32-9.
- Baur JA, Sinclair DA. Therapeutic potential of resveratrol: the in vivo evidence. Nat Rev Drug Discov. 2006;5(6):493-506.
- Borriello A, Bencivenga D, Caldarelli I, Tramontano A, Borgia A, Zappia V, et al., Resveratrol: from basic studies to bedside, in Advances in Nutrition and Cancer. Springer; 2014:167-84.
- Soleas GJ, Diamandis EP, Goldberg DM. Resveratrol: a molecule whose time has come? And gone? Clin Biochem. 1997;30(2):91-113.
- Neves AR, Lucio M, Lima LC, Reis S. Resveratrol in medicinal chemistry: a critical review of its pharmacokinetics, drug-delivery, and membrane interactions. Curr Med Chem. 2012;19(11):1663-81.
- Smoliga JM, Baur JA, Hausenblas HA. Resveratrol and health–a comprehensive review of human clinical trials. Mol Nutr Food Res. 2011;55(8):1129-41.
- Szkudelski T, Szkudelska K. Anti-diabetic effects of resveratrol. Ann N Y Acad Sci. 2011;1215(1):34-9.
- Liu K, Zhou R, Wang B, Mi MT. Effect of resveratrol on glucose control and insulin sensitivity: a meta-analysis of 11 randomized controlled trials. Am J Clin Nutr. 2014;99(6):1510-9.
- 36. Palsamy P, Subramanian S. Resveratrol, a natural phytoalexin,

normalizes hyperglycemia in streptozotocin-nicotinamide induced experimental diabetic rats. Biomed Pharmacother. 2008;62(9):598-605.

- Liu K, Zhou R, Wang B, Mi MT. Effect of resveratrol on glucose control and insulin sensitivity: a meta-analysis of 11 randomized controlled trials. Am J Clin Nutr. 2014;99(6):1510-9.
- Koushki M, Dashatan NA, Meshkani R. Effect of Resveratrol Supplementation on Inflammatory Markers: A Systematic Review and Meta-analysis of Randomized Controlled Trials. Clin Ther. 2018;40(7):1180-1192.e5.
- Alarcon De La Lastra C, Villegas I. Resveratrol as an antiinflammatory and anti-aging agent: Mechanisms and clinical implications. Mol Nutr Food Res. 2005;49(5):405-30.
- Zordoky BN, Robertson IM, Dyck JR. Preclinical and clinical evidence for the role of resveratrol in the treatment of cardiovascular diseases. Biochim Biophys Acta Mol Basis Dis. 2015;1852(6):1155-77.
- Carreras A, Zhang SX, Peris E, Qiao Z, Wang Y, Almendros I et al., Effect of resveratrol on visceral white adipose tissue inflammation and insulin sensitivity in a mouse model of sleep apnea. Int J Obes (Lond). 2015;39(3):418-23.
- 42. Jeon BT, Jeong EA, Shin HJ, Lee Y, Lee DH, Kim HJ et al., Resveratrol attenuates obesity-associated peripheral and central inflammation and improves memory deficit in mice fed a high-fat diet Diabetes. Diabetes. 2012;61(6):1444-54.
- 43. Carreras A, Zhang SX, Almendros I, Wang Y, Peris E, Qiao Z et al., Resveratrol attenuates intermittent hypoxia-induced macrophage migration to visceral white adipose tissue and insulin resistance in male mice. Endocrinology. 2014 ;156 (2) :437-43.
- Nøhr MK, Dudele A, Poulsen MM, Ebbesen LH, Radko Y, Christensen LP et al., LPS-enhanced glucose-stimulated insulin secretion is normalized by resveratrol. PLoS One. 2016;11 (1).
- 45. Lv Zm, Wang Q, Chen Yh, Wang Sh, Huang Dq. Resveratrol attenuates inflammation and oxidative stress in epididymal white adipose tissue : Implications for its involvement in improving steroidogenesis in diet-induced obese mice. Mol Reprod Dev. 2015 ;82 (4) :321-8.
- Gómez-Zorita S, Fernández-Quintela A, Lasa A, Hijona E, Bujanda L, Portillo MP. Effects of resveratrol on obesityrelated inflammation markers in adipose tissue of genetically obese rats. Nutrition. 2013;29(11):1374-80.
- Ølholm J, Paulsen S, Cullberg K, Richelsen B, Pedersen SB. Anti-inflammatory effect of resveratrol on adipokine expression and secretion in human adipose tissue explants. Int J Obes (Lond). 2010;34(10):1546-53.
- Krawczyk SA, Haller JF, Ferrante T, Zoeller RA, Corkey BE. Reactive oxygen species facilitate translocation of hormone sensitive lipase to the lipid droplet during lipolysis in human differentiated adipocytes. PLoS One. 2012;7(4):e34904.
- 49. Sun Y, Li J, Xiao N, Wang M, Kou J, Qi L et al., Pharmacological activation of AMPK ameliorates perivascular adipose/endothelial dysfunction in a manner interdependent on AMPK and SIRT1. Pharmacol Res. 2014;89:19-28.
- 50. Jimenez-Gomez Y, Mattison JA, Pearson KJ, Martin-Montalvo A, Palacios HH, Sossong AM et al., Resveratrol improves adipose insulin signaling and reduces the inflammatory response in adipose tissue of rhesus monkeys on high-fat, high-sugar diet. Cell Metab. 2013;18(4):533-45.

- Lin QQ, Yan CF, Lin R, Zhang JY, Wang WR, Yang LN, et al. SIRT1 regulates TNF-α-induced expression of CD40 in 3T3-L1 adipocytes via NF-κB pathway. Cytokine. 2012;60(2):447-55.
- Zagotta I, Dimova EY, Funcke JB, Wabitsch M, Kietzmann T, Fischer-Posovszky P. Resveratrol suppresses PAI-1 gene expression in a human in vitro model of inflamed adipose tissue. Oxid Med Cell Longev. 2013;2013.
- 53. Li X, Li J, Wang L, Li A, Qiu Z, Qi LW, et al. The role of metformin and resveratrol in the prevention of hypoxiainducible factor 1α accumulation and fibrosis in hypoxic adipose tissue. Br J Pharmacol. 2016;173(12):2001-15.
- Kim S, Jin Y, Choi Y, Park T. Resveratrol exerts anti-obesity effects via mechanisms involving down-regulation of adipogenic and inflammatory processes in mice. Biochem Pharmacol. 2011;81(11):1343-51.
- 55. Ahn J, Lee H, Kim S, Ha T. Resveratrol inhibits TNFα-induced changes of adipokines in 3T3-L1 adipocytes. Biochem Biophys Res Commun. 2007;364(4):972-7.
- 56. Yen GC, Chen YC, Chang WT. Effects of polyphenolic compounds on tumor necrosis factor-α (TNF-α)-induced changes of adipokines and oxidative stress in 3T3-L1 adipocytes. J Agric Food Chem. 2010;59(2):546-51.
- Zagotta I, Dimova EY, Debatin KM, Wabitsch M, Kietzmann T, Fischer-Posovszky P. Obesity and inflammation: reduced cytokine expression due to resveratrol in a human in vitro model of inflamed adipose tissue. Front Pharmacol. 2015;6.
- Zhu J, Yong W, Wu X, Yu Y, Lv J, Liu C et al., Antiinflammatory effect of resveratrol on TNF-α-induced MCP-1 expression in adipocytes. Biochem Biophys Res Commun. 2008 ;369 (2) :471-7.
- 59. Chuang CC, Martinez K, Xie G, Kennedy A, Bumrungpert A, Overman A et al., Quercetin is equally or more effective than resveratrol in attenuating tumor necrosis factor-α– mediated inflammation and insulin resistance in primary human adipocytes. Am J Clin Nutr. 2010;92 (6):1511-21.
- Lee HJ, Lim Y, Yang SJ. Involvement of resveratrol in crosstalk between adipokine adiponectin and hepatokine fetuin-A in vivo and in vitro. J Nutr Biochem. 2015 ;26 (11) :1254-60.
- Lettieri Barbato D, Tatulli G, Aquilano K, Ciriolo MR. Inhibition of age-related cytokines production by ATGL: a mechanism linked to the anti-inflammatory effect of resveratrol. Mediators Inflamm. 2014;2014.
- 62. Li X, Zhang S, Li J, Liu K, Huang F, Liu B. Metformin and resveratrol inhibit Drp1-mediated mitochondrial fission and prevent ER stress-associated NLRP3 inflammasome activation in the adipose tissue of diabetic mice. Mol Cell Endocrinol. 2016.
- Meshkani R, Adeli K. Hepatic insulin resistance, metabolic syndrome and cardiovascular disease. Clin Biochem. 2009;42(13):1331-46.
- Zamani-Garmsiri F, Hashemnia SMR, Shabani M, Bagherieh M, Emamgholipour S, Meshkani R. Combination of metformin and genistein alleviates non-alcoholic fatty liver disease in high-fat diet-fed mice. J Nutr Biochem. 2021;87:108505.
- Yang SJ, Lim Y. Resveratrol ameliorates hepatic metaflammation and inhibits NLRP3 inflammasome activation. Metabolism. 2014;63(5):693-701.
- 66. Faghihzadeh F, Adibi P, Rafiei R, Hekmatdoost A. Resveratrol supplementation improves inflammatory biomarkers in

patients with nonalcoholic fatty liver disease. Nutr Res. 2014;34(10):837-43.

- 67. Chen S, Zhao X, Ran L, Wan J, Wang X, Qin Y et al., Resveratrol improves insulin resistance, glucose and lipid metabolism in patients with non-alcoholic fatty liver disease: a randomized controlled trial. Dig Liver Dis. 2015 ;47 (3) :226-32.
- Nishikawa K, Iwaya K, Kinoshita M, Fujiwara Y, Akao M, Sonoda M et al., Resveratrol increases CD68+ Kupffer cells colocalized with adipose differentiation-related protein and ameliorates high-fat-diet-induced fatty liver in mice. Mol Nutr Food Res. 2015;59(6):1155-70.
- Wan J, Benkdane M, Teixeira-Clerc F, Bonnafous S, Louvet A, Lafdil F et al., M2 Kupffer cells promote M1 Kupffer cell apoptosis: a protective mechanism against alcoholic and nonalcoholic fatty liver disease. Hepatology. 2014;59(1):130-42.
- Chan CC, Cheng LY, Lin CL, Huang YH, Lin HC, Lee FY. The protective role of natural phytoalexin resveratrol on inflammation, fibrosis and regeneration in cholestatic liver injury. Mol Nutr Food Res. 2011;55(12):1841-9.
- Chan CC, Lee KC, Huang YH, Chou CK, Lin HC, Lee FY. Regulation by resveratrol of the cellular factors mediating liver damage and regeneration after acute toxic liver injury. J Gastroenterol Hepatol. 2014;29(3):603-13.
- Kessoku T, Imajo K, Honda Y, Kato T, Ogawa Y, Tomeno W et al., Resveratrol ameliorates fibrosis and inflammation in a mouse model of nonalcoholic steatohepatitis. Sci Rep. 2016;6.
- Jeong JH, Lee YR, Park HG, Lee WL. The effects of either resveratrol or exercise on macrophage infiltration and switching from M1 to M2 in high fat diet mice. J Exerc Nutr Biochem. 2015;19(2):65.
- 74. Pan QR, Ren YL, Liu WX, Hu YJ, Zheng JS, Xu Y et al., Resveratrol prevents hepatic steatosis and endoplasmic reticulum stress and regulates the expression of genes involved in lipid metabolism, insulin resistance, and inflammation in rats. Nutr Res. 2015;35(7):576-84.
- Chang CC, Chang CY, Huang JP, Hung LM. Effect of resveratrol on oxidative and inflammatory stress in liver and spleen of streptozotocin-induced type 1 diabetic rats. Chin J Physiol. 2012;55(3):192-201.
- Albertoni G, Schor N. Resveratrol plays important role in protective mechanisms in renal disease-mini-review. J Bras Nefrol. 2015;37(1):106-114.
- Tung BT, Rodríguez-Bies E, Talero E, Gamero-Estévez E, Motilva V, Navas P, et al. Anti-inflammatory effect of resveratrol in old mice liver. Exp Gerontol. 2015;64:1-7.
- Russo GT, Giorda CB, Cercone S, Nicolucci A, Cucinotta D. Factors Associated with Beta-Cell Dysfunction in Type 2 Diabetes: The BETADECLINE Study. PLoS One. 2014;9(10).
- Ehses JA, Perren A, Eppler E, Ribaux P, Pospisilik JA, Maor-Cahn R, et al. Increased number of islet-associated macrophages in type 2 diabetes. Diabetes. 2007;56(9):2356-2370.
- Li ZD, Ma QY, Wang CA. Effect of resveratrol on pancreatic oxygen free radicals in rats with severe acute pancreatitis. World J Gastroenterol. 2006;12(1):137.
- Tsang SW, Guan YF, Wang J, Bian ZX, Zhang HJ. Inhibition of pancreatic oxidative damage by stilbene derivative dihydro-resveratrol: implication for treatment of acute

pancreatitis. Sci Rep. 2016;6.

- Lee S-M, Yang H, Tartar D, Gao B, Luo X, Ye S, et al. Prevention and treatment of diabetes with resveratrol in a non-obese mouse model of type 1 diabetes. Diabetologia. 2011;54(5):1136-46.
- Khalafani Z, Zamani-Garmsiri F, Panahi G, Meshkani R. Metformin-chlorogenic acid combination reduces skeletal muscle inflammation in c57BL/6 mice on high-fat diets. Mol Biol Rep. 2023;50(3):2581-9.
- Khodabandehloo H, Gorgani-Firuzjaee S, Panahi G, Meshkani R. Molecular and cellular mechanisms linking inflammation to insulin resistance and beta-cell dysfunction. Transl Res. 2016;167(1):228-56.
- Timmers S, Konings E, Bilet L, Houtkooper RH, van de Weijer T, Goossens GH, et al. Calorie restriction-like effects of 30 days of resveratrol supplementation on energy metabolism and metabolic profile in obese humans. Cell Metab. 2011;14(5):612-22.
- Centeno-Baez C, Dallaire P, Marette A. Resveratrol inhibition of inducible nitric oxide synthase in skeletal muscle involves AMPK but not SIRT1. Am J Physiol Endocrinol Metab. 2011;301(5):E922-30.
- Sadeghi A, Seyyed Ebrahimi SS, Golestani A, Meshkani R. Resveratrol Ameliorates Palmitate-Induced Inflammation in Skeletal Muscle Cells by Attenuating Oxidative Stress and JNK/NF-kappaB Pathway in a SIRT1-Independent Mechanism. J Cell Biochem. 2017;118(9):2654-63.
- Olesen J, Gliemann L, Bienso R, Schmidt J, Hellsten Y, Pilegaard H. Exercise training, but not resveratrol, improves metabolic and inflammatory status in skeletal muscle of aged men. J Physiol (Lond). 2014;592(8):1873-86.
- Jeong JH, Park HG, Lee YR, Lee WL. Moderate exercise training is more effective than resveratrol supplementation for ameliorating lipid metabolic complication in skeletal muscle of high fat diet-induced obese mice. J Exerc Nutr Biochem (JENB). 2015;19(2):131-7.
- van der Heijden RA, Bijzet J, Meijers WC,Yakala GK,Kleemann R,Nguyen TQ,et al. Obesity-induced chronic inflammation in high fat diet challenged C57BL/6J mice is associated with acceleration of age-dependent renal amyloidosis. Sci Rep. 2015 ;5.
- 91. Tang J, Yan H, Zhuang S. Inflammation and oxidative stress in obesity-related glomerulopathy. Int J Nephrol. 2012;2012.
- Guebre-Egziabher F, Kalbacher E, Fouque D. [Insulin resistance and inflammation in chronic kidney diseases]. Nephrol Ther. 2009;5:S346-52.
- Mora C, Navarro JF. Inflammation and diabetic nephropathy. Curr Diab Rep. 2006;6(6):463-8.
- Lim AK, Tesch GH. Inflammation in diabetic nephropathy. Mediators Inflamm. 2012;2012.
- Biswas SK, Lopes de Faria J. Hypertension induces oxidative stress but not macrophage infiltration in the kidney in the early stage of experimental diabetes mellitus. Am J Nephrol. 2006;26(5):415-22.
- Lee VWS, Wang YM, Wang Y, Zheng D, Polhill T, Cao Q, et al. Regulatory immune cells in kidney disease. Am J Physiol Renal Physiol. 2008;295(2):F335-42.
- Hartner A, Veelken R, Wittmann M, Cordasic N, Hilgers KF. Effects of diabetes and hypertension on macrophage infiltration and matrix expansion in the rat kidney. BMC Nephrol. 2005;6(1):1.
- 98. Wada J, Makino H. Innate immunity in diabetes and diabetic

nephropathy. Nat Rev Nephrol. 2015.

- Roubicek T, Bartlova M, Krajickova J, Haluzikova D, Mraz M, Lacinova Z, et al. Increased production of proinflammatory cytokines in adipose tissue of patients with end-stage renal disease. Nutrition (Burbank). 2009;25(7):762-8.
- 100. Egido J, Gomez-Chiarri M, Ortiz A, Bustos C, Alonso J, Gomez-Guerrero C, et al. Role of tumor necrosis factor-alpha in the pathogenesis of glomerular diseases. Kidney Int Suppl (2011). 1993;39:S59-64.
- 101. Patel NS, Chatterjee PK, Di Paola R, Mazzon E, Britti D, De Sarro A, et al. Endogenous interleukin-6 enhances the renal injury, dysfunction, and inflammation caused by ischemia/ reperfusion. J Pharmacol Exp Ther. 2005;312(3):1170-8.
- 102. Lan HY, Bacher M, Yang N, Mu W, Nikolic-Paterson DJ, Metz C, et al. The pathogenic role of macrophage migration inhibitory factor in immunologically induced kidney disease in the rat. J Exp Med. 1997;185(8):1455-66.
- 103. Zhang L, Pang S, Deng B, Qian L, Chen J, Zou J, et al. High glucose induces renal mesangial cell proliferation and fibronectin expression through JNK/NF-kappaB/NADPH oxidase/ROS pathway, which is inhibited by resveratrol. Int J Biochem Cell Biol. 2012;44(4):629-38.
- 104. Palsamy P, Subramanian S. Resveratrol protects diabetic kidney by attenuating hyperglycemia-mediated oxidative stress and renal inflammatory cytokines via Nrf2–Keap1 signaling. Biochim Biophys Acta Mol Basis Dis. 2011;1812(7):719-31.
- 105. Kim MY, Lim JH, Youn HH, Hong YA, Yang KS, Park HS, et al. Resveratrol prevents renal lipotoxicity and inhibits mesangial cell glucotoxicity in a manner dependent on the AMPK-SIRT1-PGC1alpha axis in db/db mice. Diabetologia. 2013;56(1):204-17.
- Sharma S, Anjaneyulu M, Kulkarni S, Chopra K. Resveratrol attenuates diabetic nephropathy in rats. Pharmacology (Basel). 2006;76(2):69-75.
- 107. Zhou Y, Lin S, Zhang L, Li Y. Resveratrol prevents renal lipotoxicity in high-fat diet-treated mouse model through regulating PPAR-alpha pathway. Mol Cell Biochem. 2016;411(1-2):143-50.
- 108. Wu M, Gu J, Mei S, Xu D, Jing Y,Yao Q,et al. Resveratrol delays polycystic kidney disease progression through attenuation of nuclear factor κB-induced inflammation. Nephrol Dial Transplant. 2016 ;gfw058.
- 109. Kolgazi M, Şener G, Çetinel Ş, Gedik N, Alican İ. Resveratrol reduces renal and lung injury caused by sepsis in rats. J Surg Res. 2006;134 (2):315-21.
- 110. Şener G, Tuğtepe H, Yüksel M, Çetinel Ş, Gedik N, Yeğen BÇ. Resveratrol improves ischemia/reperfusion-induced oxidative renal injury in rats. Arch Med Res. 2006 ;37 (7) :822-9.
- 111. Chen L, Yang S, Zumbrun EE, Guan H, Nagarkatti PS, Nagarkatti M. Resveratrol attenuates lipopolysaccharideinduced acute kidney injury by suppressing inflammation driven by macrophages. Mol Nutr Food Res. 2015;59(5):853-64.
- 112. Castilla P, Davalos A, Teruel JL, Cerrato F, Fernandez-Lucas M, Merino JL, et al. Comparative effects of dietary supplementation with red grape juice and vitamin E on production of superoxide by circulating neutrophil NADPH oxidase in hemodialysis patients. Am J Clin Nutr. 2008;87(4):1053-61.
- Castilla P, Echarri R, Davalos A, Cerrato F, Ortega H, Teruel JL, et al. Concentrated red grape juice exerts antioxidant, hypolipidemic, and antiinflammatory effects in both

hemodialysis patients and healthy subjects. Am J Clin Nutr. 2006;84(1):252-62.

- 114. Laslett LJ, Alagona P Jr., Clark BA 3rd, Drozda JP Jr., Saldivar F, Wilson SR, et al. The worldwide environment of cardiovascular disease: prevalence, diagnosis, therapy, and policy issues: a report from the American College of Cardiology. J Am Coll Cardiol. 2012;60(25 Suppl):S1-49.
- 115. Mohassel Yaser SA, Mostafae Shayan MTG. Assessing the Possible Association between Polymorphism of C677T MTHFR with Preeclampsia Risk: A Systematic Review and Bayesian Hierarchical Meta-Analysis. Acta Biochim Iranica. 2023;1(1):3-11.
- 116. Libby P, Ridker PM, Maseri A. Inflammation and atherosclerosis. Circulation. 2002;105(9):1135-43.
- 117. Wakabayashi I, Takeda Y. Inhibitory effects of resveratrol on MCP-1, IL-6, and IL-8 production in human coronary artery smooth muscle cells. Naunyn Schmiedebergs Arch Pharmacol. 2013 ;386 (9) :835-9.
- 118. Zhong M, Cheng GF, Wang WJ, Guo Y, Zhu XY, Zhang JT. Inhibitory effect of resveratrol on interleukin 6 release by stimulated peritoneal macrophages of mice. Phytomedicine. 1999;6 (2):79-84.
- 119. Song R, Li WQ, Dou JL, Li L, Hu YJ, Guo JZ, et al. [Resveratrol reduces inflammatory cytokines via inhibiting nuclear factor-kappaB and mitogen-activated protein kinase signal pathway in a rabbit atherosclerosis model]. Zhonghua Xin Xue Guan Bing Za Zhi. 2013 ;41 (10) :866-9.
- 120. Pervaiz S. Resveratrol: from grapevines to mammalian biology. Faseb j. 2003 ;17 (14) :1975-85.
- 121. Deng YH, Alex D, Huang HQ, Wang N, Yu N, Wang YT, et al. Inhibition of TNF-alpha-mediated endothelial cellmonocyte cell adhesion and adhesion molecules expression by the resveratrol derivative, trans-3,5,4'-trimethoxystilbene. Phytother Res. 2011;25(3):451-7.
- 122. Kim SW, Kim CE, Kim MH. Flavonoids inhibit high glucoseinduced up-regulation of ICAM-1 via the p38 MAPK pathway in human vein endothelial cells. Biochem Biophys Res Commun. 2011;415(4):602-7.
- 123. Scoditti E, Calabriso N, Massaro M, Pellegrino M, Storelli C, Martines G, et al. Mediterranean diet polyphenols reduce inflammatory angiogenesis through MMP-9 and COX-2 inhibition in human vascular endothelial cells: a potentially protective mechanism in atherosclerotic vascular disease and cancer. Arch Biochem Biophys. 2012;527(2):81-9.
- 124. Buttari B, Profumo E, Segoni L, D'Arcangelo D, Rossi S, Facchiano F, et al. Resveratrol counteracts inflammation in human M1 and M2 macrophages upon challenge with 7-oxo-cholesterol: potential therapeutic implications in atherosclerosis. Oxid Med Cell Longev. 2014;2014:257543.
- 125. Csiszar A, Labinskyy N, Podlutsky A, Kaminski PM,Wolin MS, Zhang C, et al. Vasoprotective effects of resveratrol and SIRT1: attenuation of cigarette smoke-induced oxidative stress and proinflammatory phenotypic alterations. Am J Physiol Heart Circ Physiol. 2008 ;294 (6) :H2721-35.
- 126. Latruffe N, Lancon A, Frazzi R, Aires V, Delmas D, Michaille JJ, et al. Exploring new ways of regulation by resveratrol involving miRNAs, with emphasis on inflammation. Ann N Y Acad Sci. 2015;1348 (1):97-106.
- 127. Li J, Xie C, Zhuang J, Li H, Yao Y, Shao C, et al. Resveratrol attenuates inflammation in the rat heart subjected to ischemiareperfusion: role of the TLR4/NF-kappaB signaling pathway. Mol Med Rep. 2015 ;11 (2) :1120-6.

- 128. Dong W, Yang R, Yang J, Yang J, Ding J, Wu H, et al. Resveratrol pretreatment protects rat hearts from ischemia/ reperfusion injury partly via a NALP3 inflammasome pathway. Int J Clin Exp Pathol. 2015 ;8 (8) :8731-41.
- 129. Cheng L, Jin Z, Zhao R, Ren K, Deng C, Yu S. Resveratrol attenuates inflammation and oxidative stress induced by myocardial ischemia-reperfusion injury: role of Nrf2/ARE pathway. Int J Clin Exp Med. 2015 ;8 (7) :10420.
- McMurray JJ, Pfeffer MA. Heart failure. Lancet. 2005 ;365 (9474) :1877-89.
- Gupta PK, DiPette DJ, Supowit SC. Protective effect of resveratrol against pressure overload-induced heart failure. Food Sci Nutr. 2014;2(3):218-29.
- Yoshida Y, Shioi T, Izumi T. Resveratrol ameliorates experimental autoimmune myocarditis. Circ J. 2007;71(3):397-404.
- 133. Tomé-Carneiro J, Larrosa M, Yánez-Gascón MJ, Dávalos A, Gil-Zamorano J, Gonzálvez M, et al. One-year supplementation with a grape extract containing resveratrol modulates inflammatory-related microRNAs and cytokines expression in peripheral blood mononuclear cells of type 2 diabetes and hypertensive patients with coronary artery disease. Pharmacol Res. 2013;72:69-82.
- 134. Tomé-Carneiro J, Gonzálvez M, Larrosa M, Yáñez-Gascón MJ, García-Almagro FJ, Ruiz-Ros JA, et al. Grape resveratrol increases serum adiponectin and downregulates inflammatory genes in peripheral blood mononuclear cells: a triple-blind, placebo-controlled, one-year clinical trial in patients with stable coronary artery disease. Cardiovasc Drugs Ther. 2013;27(1):37-48.
- 135. Tomé-Carneiro J, Larrosa M, Yánez-Gascón MJ, Dávalos A, Gil-Zamorano J, Gonzálvez M, et al. One-year supplementation with a grape extract containing resveratrol modulates inflammatory-related microRNAs and cytokines expression in peripheral blood mononuclear cells of type 2 diabetes and hypertensive patients with coronary artery disease. Pharmacol Res. 2013;72:69-82.
- 136. Tomé-Carneiro J, Gonzálvez M, Larrosa M, Yáñez-Gascón MJ, García-Almagro FJ, Ruiz-Ros JA, et al. Grape resveratrol increases serum adiponectin and downregulates inflammatory genes in peripheral blood mononuclear cells: a triple-blind, placebo-controlled, one-year clinical trial in patients with stable coronary artery disease. Cardiovasc Drugs Ther. 2013;27(1):37-48.
- 137. Tomé-Carneiro J, Gonzálvez M, Larrosa M, Yáñez-Gascón MJ, García-Almagro FJ, Ruiz-Ros JA, et al. One-year consumption of a grape nutraceutical containing resveratrol improves the inflammatory and fibrinolytic status of patients in primary prevention of cardiovascular disease. Am J Cardiol. 2012;110 (3):356-63.
- 138. Magyar K, Halmosi R, Palfi A, Feher G, Czopf L, Fulop A, et al. Cardioprotection by resveratrol: a human clinical trial in patients with stable coronary artery disease. Clin Hemorheol Microcirc. 2012 ;50 (3) :179-87.
- Franco R, Fernandez-Suarez D. Alternatively activated microglia and macrophages in the central nervous system. Prog Neurobiol. 2015;131:65-86.
- 140. Minogue AM. Role of infiltrating monocytes/macrophages in acute and chronic neuroinflammation: Effects on cognition, learning and affective behaviour. Prog Neuropsychopharmacol Biol Psychiatry. 2017 ;79 (Pt A) :15-18.
- 141. Zhang F, Liu J, Shi JS. Anti-inflammatory activities of

resveratrol in the brain: role of resveratrol in microglial activation. Eur J Pharmacol. 2010;636(1):1-7.

- 142. Choi DK, Koppula S, Suk K. Inhibitors of microglial neurotoxicity: focus on natural products. Molecules. 2011;16(2):1021-43.
- 143. Fang L, Gao H, Zhang W, Zhang W, Wang Y. Resveratrol alleviates nerve injury after cerebral ischemia and reperfusion in mice by inhibiting inflammation and apoptosis. Int J Clin Exp Med. 2015;8(3):3219.
- 144. Im Jeong S, Shin JA, Cho S, Kim HW, Lee JY, Kang JL, et al. Resveratrol attenuates peripheral and brain inflammation and reduces ischemic brain injury in aged female mice. Neurobiol Aging. 2016;44:74-84.
- 145. Gatson JW, Liu MM, Abdelfattah K, Wigginton JG, Smith S, Wolf S, et al. Resveratrol decreases inflammation in the brain of mice with mild traumatic brain injury. J Trauma Acute Care Surg. 2013;74(2):470-5.
- 146. Hoda U, Agarwal NB, Vohora D, Parvez S, Raisuddin S. Resveratrol attenuates seizures by attenuating IL-1β, IL1-Ra, IL-6, and TNF-α in the hippocampus and cortex of kindled mice. Nutr Neurosci. 2016 ;1-8.
- 147. Tian X, Liu Y, Ren G, Yin L, Liang X, Geng T, et al. Resveratrol limits diabetes-associated cognitive decline in rats by preventing oxidative stress and inflammation and modulating hippocampal structural synaptic plasticity. Brain Res. 2016 ;1650 :1-9.
- 148. Hoda U, Agarwal NB, Vohora D, Parvez S, Raisuddin S. Resveratrol attenuates seizures by attenuating IL-1β, IL1-Ra, IL-6, and TNF-α in the hippocampus and cortex of kindled mice. Nutr Neurosci. 2017 ;20 (9) :497-504.
- 149. Mishra V, Shuai B, Kodali M, Shetty GA, Hattiangady B, Rao X, et al. Resveratrol treatment after status epilepticus restrains neurodegeneration and abnormal neurogenesis with suppression of oxidative stress and inflammation. Sci Rep. 2015;5:17807.
- 150. Ge L, Liu L, Liu H, Liu S, Xue H, Wang X, et al. Resveratrol abrogates lipopolysaccharide-induced depressive-like behavior, neuroinflammatory response, and CREB/BDNF signaling in mice. Eur J Pharmacol. 2015 ;768 :49-57.
- 151. Tiwari V, Chopra K. Resveratrol prevents alcohol-induced cognitive deficits and brain damage by blocking inflammatory signaling and cell death cascade in neonatal rat brain. J Neurochem. 2011;117(4):678-90.
- Choi DK, Koppula S, Suk K. Inhibitors of microglial neurotoxicity: focus on natural products. Molecules. 2011;16(2):1021-43.
- 153. Fang L, Gao H, Zhang W, Zhang W, Wang Y. Resveratrol alleviates nerve injury after cerebral ischemia and reperfusion in mice by inhibiting inflammation and apoptosis. Int J Clin Exp Med. 2015;8(3):3219.
- 154. Zhong L-M, Zong Y, Sun L, Guo J-Z, Zhang W, He Y, et al. Resveratrol inhibits inflammatory responses via the mammalian target of rapamycin signaling pathway in cultured LPS-stimulated microglial cells. PLoS One. 2012;7(2):e32195.
- 155. Bureau G, Longpré F, Martinoli MG. Resveratrol and quercetin, two natural polyphenols, reduce apoptotic neuronal cell death induced by neuroinflammation. J Neurosci Res. 2008;86(2):403-10.
- 156. Wyss-Coray T, Mucke L. Inflammation in neurodegenerative disease-a double-edged sword. Neuron. 2002;35(3):419-32.
- 157. Rudge JS, Morrissey D, Lindsay RM, Pasnikowski EM.

Regulation of ciliary neurotrophic factor in cultured rat hippocampal astrocytes. Eur J Neurosci. 1994 ;6 (2) :218-29.

- 158. Kim YA, Kim GY, Park KY, Choi YH. Resveratrol inhibits nitric oxide and prostaglandin E2 production by lipopolysaccharide-activated C6 microglia. J Med Food. 2007 ;10 (2) :218-24.
- 159. White KA, Hutton SR, Weimer JM, Sheridan PA. Dietinduced obesity prolongs neuroinflammation and recruits CCR2+ monocytes to the brain following herpes simplex virus (HSV)-1 latency in mice. Brain Behav Immun. 2016.
- 160. Beilharz JE, Maniam J, Morris MJ. Short-term exposure to a diet high in fat and sugar, or liquid sugar, selectively impairs hippocampal-dependent memory with differential impacts on inflammation. Behav Brain Res. 2016 ;306 :1-7.
- 161. Spagnuolo MS, Mollica MP, Maresca B, Cavaliere G, Cefaliello C, Trinchese G, et al. High Fat Diet and Inflammation–Modulation of Haptoglobin Level in Rat Brain. Front Cell Neurosci. 2015;9.
- 162. Miller AA, Spencer SJ. Obesity and neuroinflammation: a pathway to cognitive impairment. Brain Behav Immun. 2014;42:10-21.
- 163. Cai D. Neuroinflammation and neurodegeneration in overnutrition-induced diseases. Trends Endocrinol Metab. 2013;24(1):40-7.
- 164. Kumar A, Sharma SS. NF-kappaB inhibitory action of resveratrol: a probable mechanism of neuroprotection in experimental diabetic neuropathy. Biochem Biophys Res Commun. 2010;394(2):360-5.
- Vinik AI, Mehrabyan A. Diabetic neuropathies. Med Clin North Am. 2004;88(4):947-99, xi.
- Ramadori G, Gautron L, Fujikawa T, Vianna CR, Elmquist JK, Coppari R. Central administration of resveratrol improves diet-induced diabetes. Endocrinology. 2009;150(12):5326-33.
- 167. Thomas J, Garg ML, Smith DW. Dietary resveratrol supplementation normalizes gene expression in the hippocampus of streptozotocin-induced diabetic C57Bl/6 mice. J Nutr Biochem. 2014;25(3):313-8.
- 168. Kumar A, Sharma SS. NF-κB inhibitory action of resveratrol: a probable mechanism of neuroprotection in experimental diabetic neuropathy. Biochem Biophys Res Commun. 2010 ;394 (2) :360-5.
- 169. Kumar A, Kaundal RK, Iyer S, Sharma SS. Effects of resveratrol on nerve functions, oxidative stress and DNA fragmentation in experimental diabetic neuropathy. Life Sci. 2007;80 (13):1236-44.
- 170. Sharma SS, Kumar A, Arora M, Kaundal RK. Neuroprotective potential of combination of resveratrol and 4-amino 1, 8 naphthalimide : focus on functional, sensorimotor and biochemical changes. Free Radic Res. 2009 ;43 (4) :400-8.
- Anekonda TS. Resveratrol—A boon for treating Alzheimer's disease? Brain Res Rev. 2006;52(2):316-26.
- 172. Zhuang H, Kim Y-S, Koehler RC, DorÉ S. Potential Mechanism by Which Resveratrol, a Red Wine Constituent, Protects Neurons. Ann N Y Acad Sci. 2003;993(1):276-86.
- 173. Clark D, Tuor UI, Thompson R, Institoris A, Kulynych A, Zhang X, et al. Protection against Recurrent Stroke with Resveratrol: Endothelial Protection. PLoS ONE. 2012;7(10):e47792.
- 174. Huang SS, Tsai MC, Chih CL, Hung LM, Tsai SK. Resveratrol reduction of infarct size in Long-Evans rats subjected to focal cerebral ischemia. Life Sci. 2001;69(9):1057-65.